

Barriers to replicating preclinical cancer biology research challenges or opportunities?

Tim Errington

Center for Open Science

<https://cos.io/>



JOHN TEMPLETON
FOUNDATION



Templeton
Religion Trust





Mission: Improve
openness, integrity, and
reproducibility of
scientific research

Norms

Communality

Open sharing

Universalism

Evaluate research on own merit

Disinterestedness

Motivated by knowledge and discovery

Organized skepticism

Consider all new evidence, even against one's prior work

Quality

Counternorms

Secrecy

Closed

Particularism

Evaluate research by reputation

Self-interestedness

Treat science as a competition

Organized dogmatism

Invest career promoting one's own theories, findings

Quantity

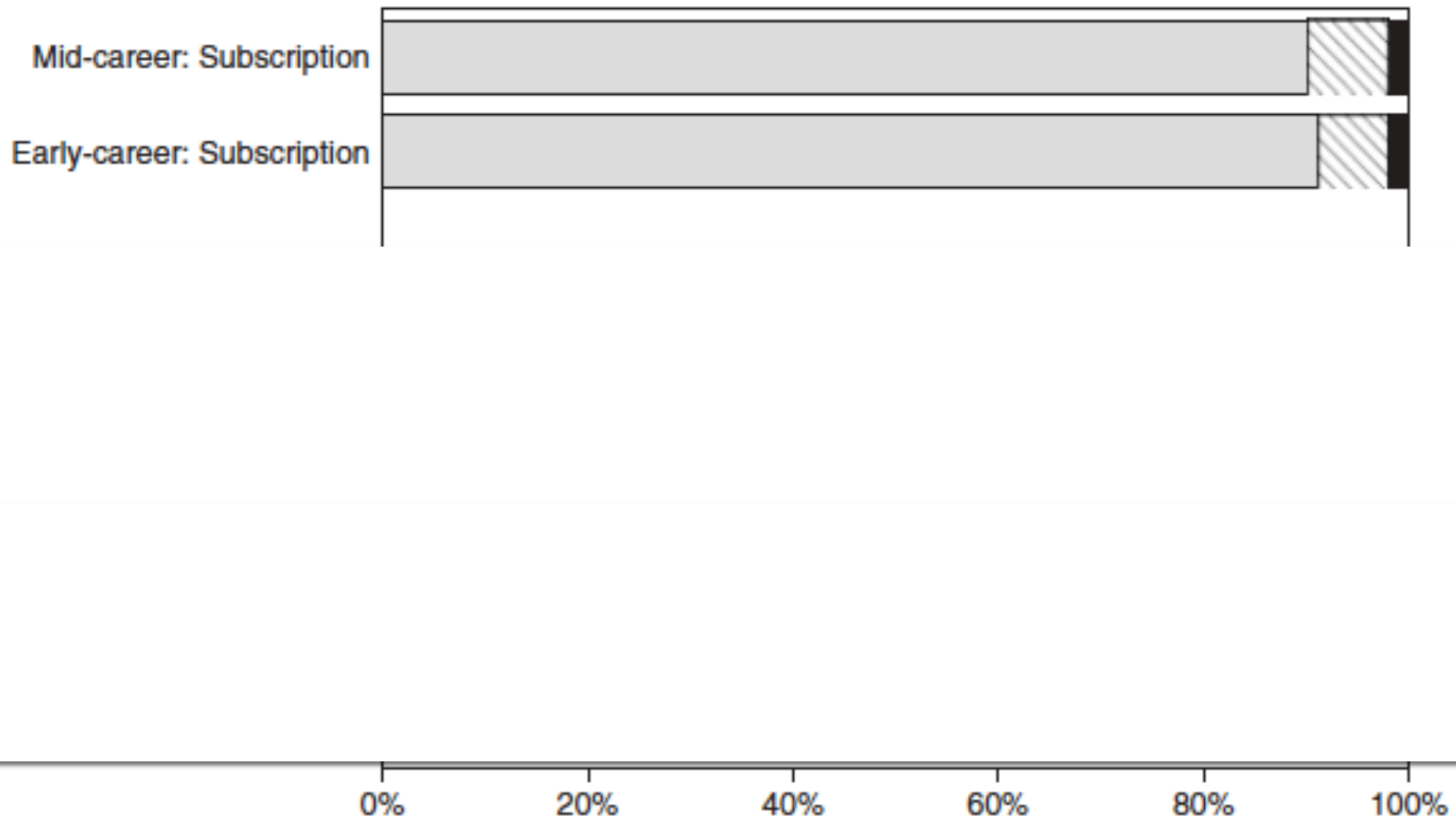


FIG. 3. Norm versus Counternorm Scores: Percent with Norm > Counternorm (dotted), Norm = Counternorm (striped), Norm < Counternorm (solid).

Barriers

1. Perceived norms (Anderson, Martinson, & DeVries, 2007)
2. Motivated reasoning (Kunda, 1990)
3. Minimal accountability (Lerner & Tetlock, 1999)
4. Concrete rewards beat abstract principles (Trope & Liberman, 2010)
5. I am busy (Me & You, 2023)
6. Incentives for individual success are focused on getting it published, not getting it right (Nosek, Spies, & Motyl, 2012)

Believe it or not: how much can we rely on published data on potential drug targets?

Florian Prinz, Thomas Schlange and Khusru Asadullah

Raise standards for preclinical cancer research

C. Glenn Begley and Lee M. Ellis propose how methods, publications and incentives must change if patients are to benefit.

Believe it or not: how much can we rely on published data on potential drug targets?

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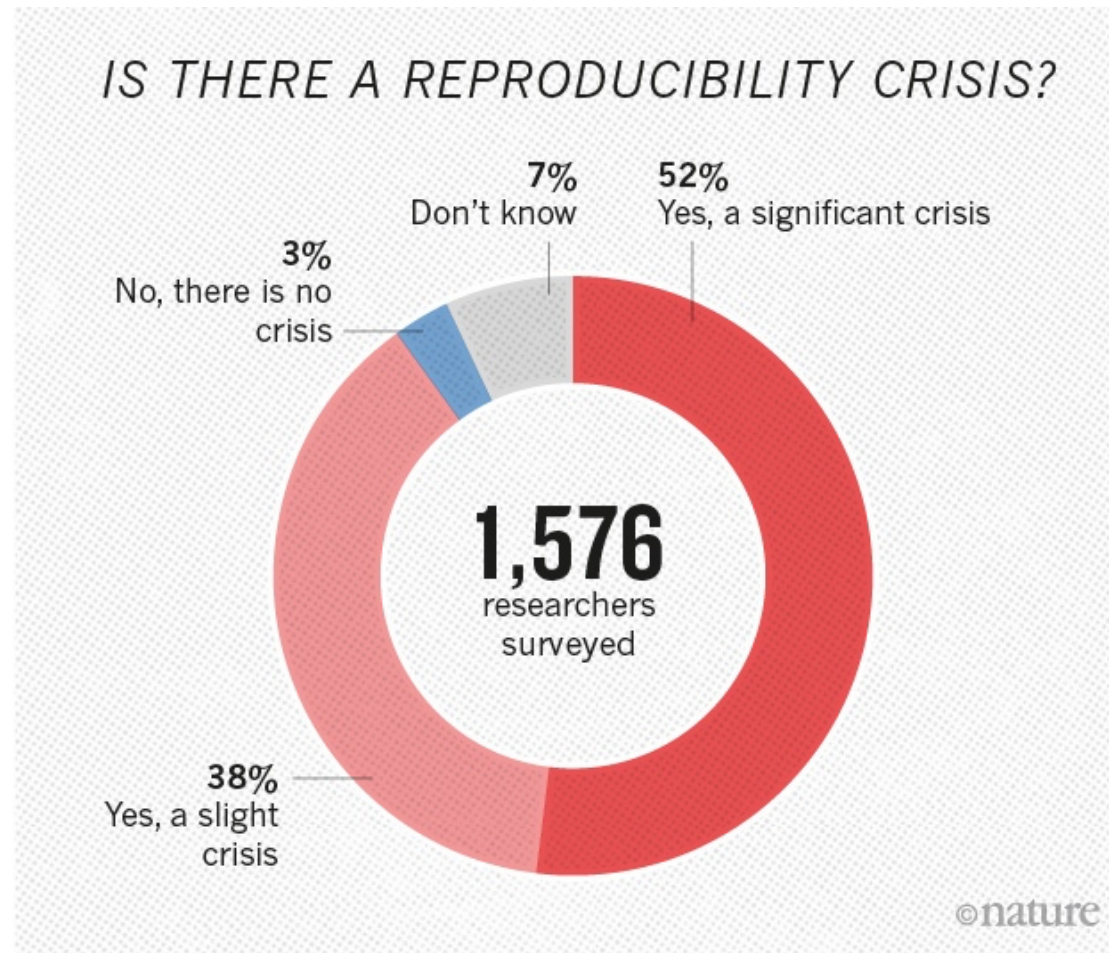
We received input from 23 scientists (heads of laboratories) and collected data from 67 projects, most of them (47) from the field of oncology. This analysis revealed that only in ~20–25% of the projects were the relevant published data completely in line with our in-house findings (FIG. 1c). In almost two-thirds

Raise standards for preclinical cancer research

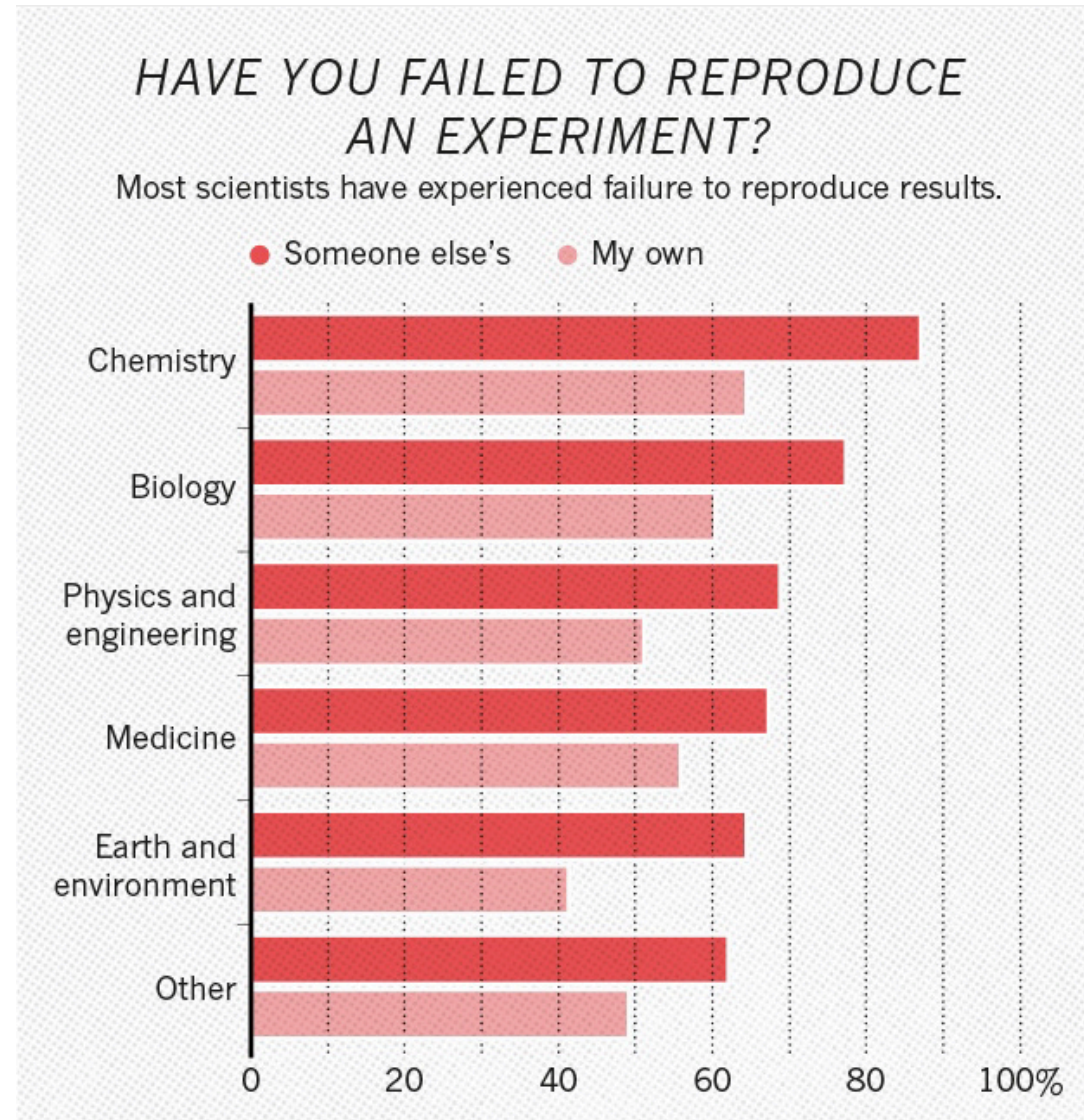
C. Glenn Begley and Lee M. Ellis propose how methods, publications and incentives must change if patients are to benefit.

tive clinical uses for existing therapeutics. Nevertheless, scientific findings were confirmed in only 6 (11%) cases. Even knowing the limitations of preclinical research, this was a shocking result.

- Nature survey of 1,576 researchers



Motivation

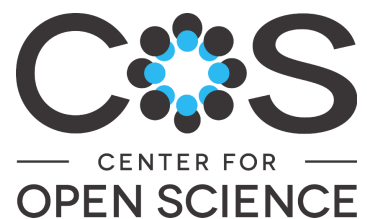


Approach



REPRODUCIBILITY PROJECT Cancer Biology

The Reproducibility Project: Cancer Biology is a collaboration between the [Center for Open Science](#) and [Science Exchange](#) to independently replicate selected results from a substantial number of [high-profile papers](#) in the field of cancer biology. For each paper a Registered Report detailing the proposed experimental designs and protocols for the replications is peer reviewed and published prior to data collection. The results of these experiments will be published in a Replication Study. [The project](#) will provide evidence about reproducibility in cancer biology, and an opportunity to identify factors that influence reproducibility more generally.



Approach

Systematic sampling of 53 high-impact preclinical cancer biology papers

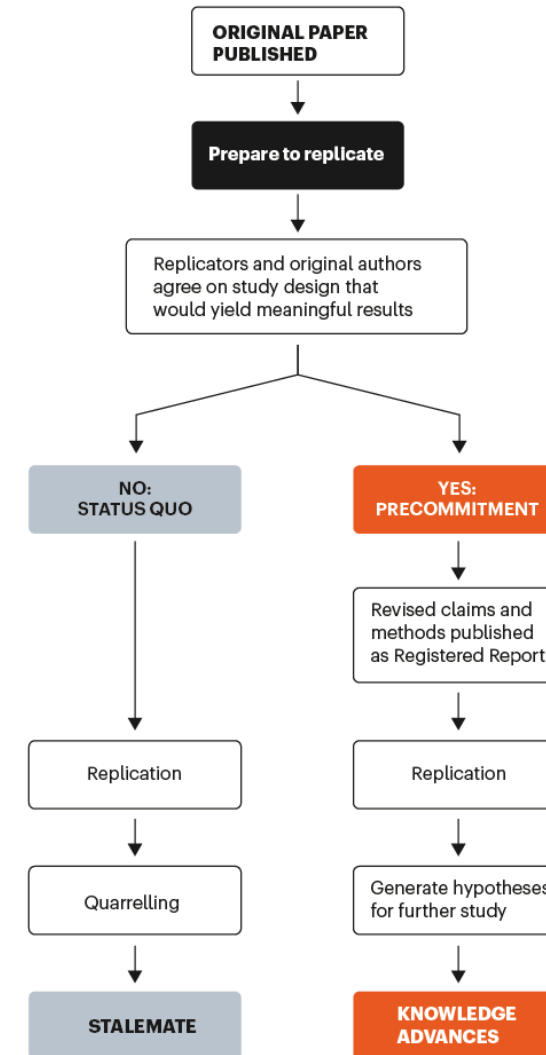
- Papers from 2010-2012
- Excluded genomics, proteomics, high-throughput assays

Primary Outcomes

- Summary of process challenges for testing replicability
- Meta-analytic summary of statistical outcomes

ROUTES TO REPLICATION

Precommitment rewards authors for providing clear, testable claims and helps to advance knowledge.



Approach

1. Identify key conclusions from the paper and a subset of data from the paper to be replicated



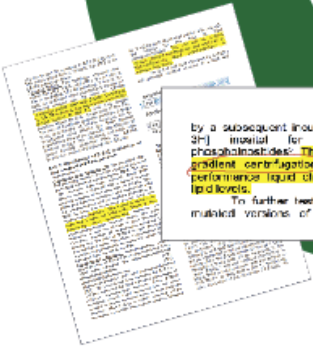
5. Perform experiments, *in vitro* and *in vivo*, according to protocol from registered report.



4. Compile obtained information into organized registered report that is published at eLife and undergoes a review process, open to commentary from study authors.



2. Obtain the materials and methods from the key figures of the paper and annotate them to determine the holes in the protocols that would influence the success of replication



6. Publish findings in an open access Replication Study



3. Contact laboratories to get them to fill in the holes. This often involves searching through old research notebooks, connecting with study authors that have since moved on, and interfacing with collaborators.



by a subsequent incubation period with myo-[2-³H]-lipid material for ~ one hour in label phospholipids. The PM was isolated through gradient centrifugation and analyzed by high-performance liquid chromatography to measure lipid levels.
To further test lipid transport by E-Syt2, mutated versions of E-Syt2 with hydrophobic



FEATURE ARTICLE



RESEARCH ARTICLE



REPRODUCIBILITY IN CANCER BIOLOGY

Challenges for assessing replicability in preclinical cancer biology

Abstract We conducted the [Reproducibility Project: Cancer Biology](#) to investigate the replicability of preclinical research in cancer biology. The initial aim of the project was to repeat 193 experiments from 53 high-impact papers, using an approach in which the experimental protocols and plans for data analysis had to be peer reviewed and accepted for publication before experimental work could begin. However, the various barriers and challenges we encountered while designing and conducting the experiments meant that we were only able to repeat 50 experiments from 23 papers. Here we report these barriers and challenges. First, many original papers failed to report key descriptive and inferential statistics: the data needed to compute effect sizes and conduct power analyses was publicly accessible for just 4 of 193 experiments. Moreover, despite contacting the authors of the original papers, we were unable to obtain these data for 68% of the experiments. Second, none of the 193 experiments were described in sufficient detail in the original paper to enable us to design protocols to repeat the experiments, so we had to seek clarifications from the original authors. While authors were *extremely* or *very helpful* for 41% of experiments, they were *minimally helpful* for 9% of experiments, and *not at all helpful* (or did not respond to us) for 32% of experiments. Third, once experimental work started, 67% of the peer-reviewed protocols required modifications to complete the research and just 41% of those modifications could be implemented. Cumulatively, these three factors limited the number of experiments that could be repeated. This experience draws attention to a basic and fundamental concern about replication – it is hard to assess whether reported findings are credible.

TIMOTHY M ERRINGTON*, ALEXANDRIA DENIS†, NICOLE PERFITO‡, ELIZABETH IORNS AND BRIAN A NOSEK

Investigating the replicability of preclinical cancer biology

Timothy M Errington^{1*}, Maya Mathur², Courtney K Soderberg¹, Alexandria Denis^{1†}, Nicole Perfito^{1‡}, Elizabeth Iorns³, Brian A Nosek^{1,4}

¹Center for Open Science, Charlottesville, United States; ²Quantitative Sciences Unit, Stanford University, Stanford, United States; ³Science Exchange, Palo Alto, United States; ⁴University of Virginia, Charlottesville, United States

Abstract Replicability is an important feature of scientific research, but aspects of contemporary research culture, such as an emphasis on novelty, can make replicability seem less important than it should be. The [Reproducibility Project: Cancer Biology](#) was set up to provide evidence about the replicability of preclinical research in cancer biology by repeating selected experiments from high-impact papers. A total of 50 experiments from 23 papers were repeated, generating data about the replicability of a total of 158 effects. Most of the original effects were positive effects (136), with the rest being null effects (22). A majority of the original effect sizes were reported as numerical values (117), with the rest being reported as representative images (41). We employed seven methods to assess replicability, and some of these methods were not suitable for all the effects in our sample. One method compared effect sizes: for positive effects, the median effect size in the replications was 85% smaller than the median effect size in the original experiments, and 92% of replication effect sizes were smaller than the original. The other methods were binary – the replication was either a success or a failure – and five of these methods could be used to assess both positive and null effects when effect sizes were reported as numerical values. For positive effects, 40% of replications (39/97) succeeded according to three or more of these five methods, and for null effects 80% of replications (12/15) were successful on this basis; combining positive and null effects, the success rate was 46% (51/112). A successful replication does not definitively confirm an original finding or its theoretical interpretation. Equally, a failure to replicate does not disconfirm a finding, but it does suggest that additional investigation is needed to establish its reliability.



FEATURE ARTICLE



RESEARCH ARTICLE



REPRODUCIBILITY IN CANCER BIOLOGY

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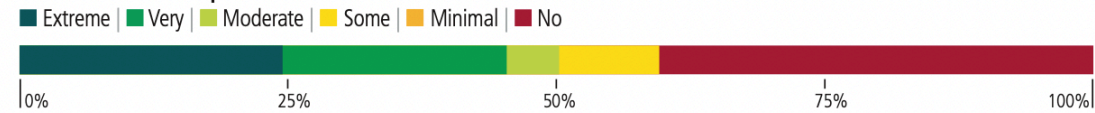
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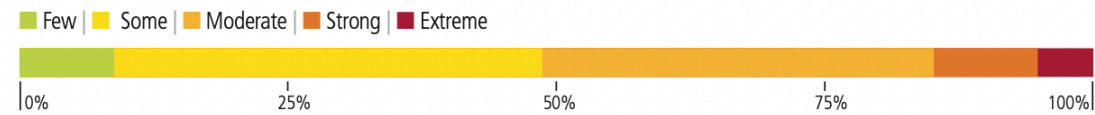
Challenges

DESIGNED
193 experiments

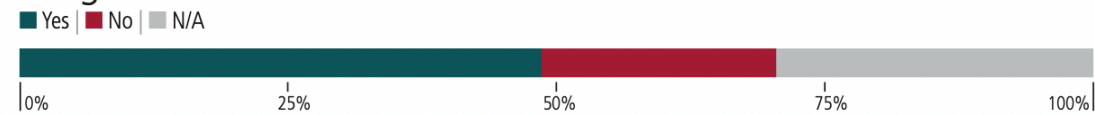
Authors helped



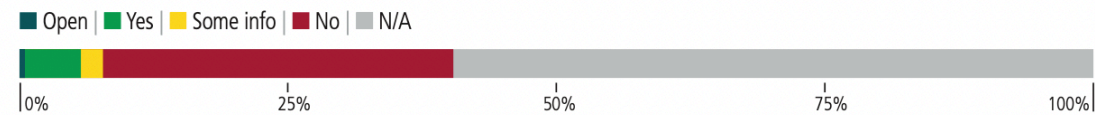
Protocol clarifications needed



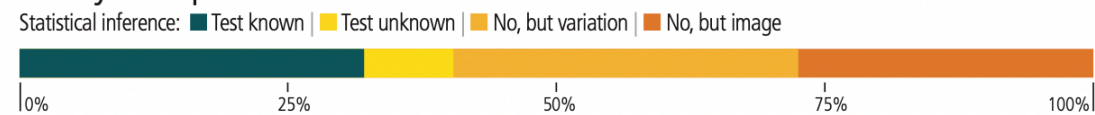
Reagents offered



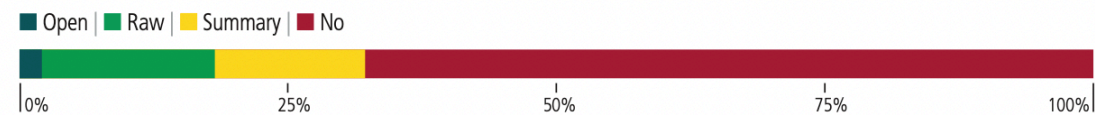
Code shared



Analysis reported



Data shared



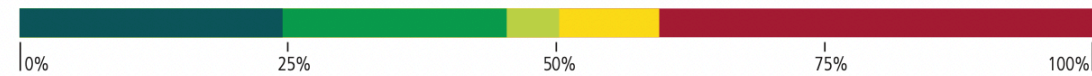
Challenges

DESIGNED
193 experiments

2% had open data; after requests 16%
shared raw data

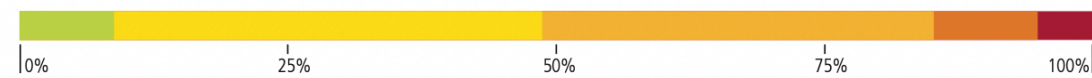
Authors helped

Extreme | Very | Moderate | Some | Minimal | No



Protocol clarifications needed

Few | Some | Moderate | Strong | Extreme



Reagents offered

Yes | No | N/A



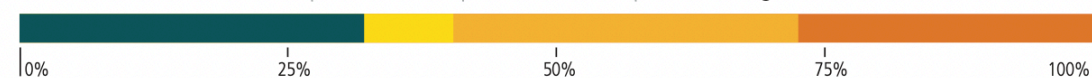
Code shared

Open | Yes | Some info | No | N/A



Analysis reported

Statistical inference: Test known | Test unknown | No, but variation | No, but image



Data shared

Open | Raw | Summary | No



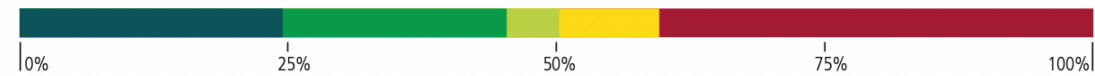
Challenges

DESIGNED
193 experiments

21% had unclear statistical tests

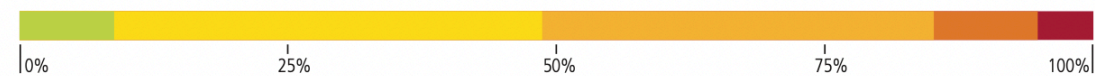
Authors helped

Extreme | Very | Moderate | Some | Minimal | No



Protocol clarifications needed

Few | Some | Moderate | Strong | Extreme



Reagents offered

Yes | No | N/A



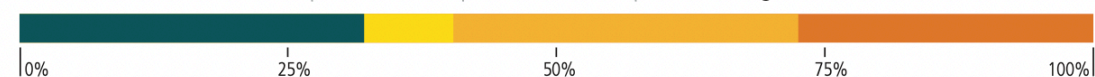
Code shared

Open | Yes | Some info | No | N/A



Analysis reported

Statistical inference: Test known | Test unknown | No, but variation | No, but image



Data shared

Open | Raw | Summary | No



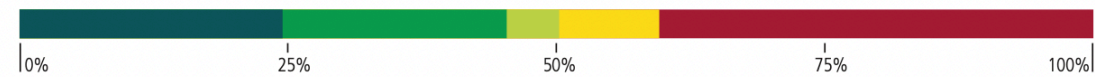
Challenges

DESIGNED
193 experiments

1% had open code; after requests 13%
shared code

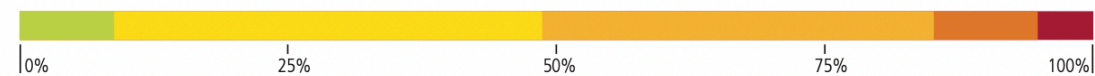
Authors helped

Extreme | Very | Moderate | Some | Minimal | No



Protocol clarifications needed

Few | Some | Moderate | Strong | Extreme



Reagents offered

Yes | No | N/A



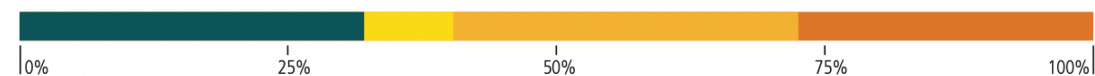
Code shared

Open | Yes | Some info | No | N/A



Analysis reported

Statistical inference: Test known | Test unknown | No, but variation | No, but image



Data shared

Open | Raw | Summary | No



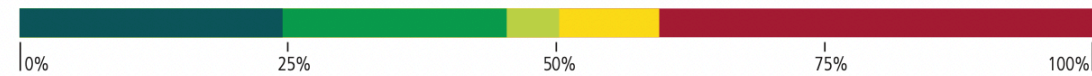
Challenges

DESIGNED
193 experiments

70% required asking for key reagents;
69% willing to share

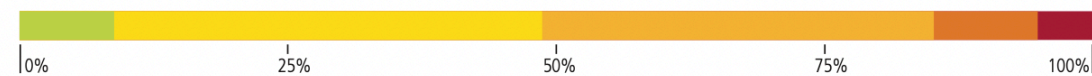
Authors helped

Extreme | Very | Moderate | Some | Minimal | No



Protocol clarifications needed

Few | Some | Moderate | Strong | Extreme



Reagents offered

Yes | No | N/A



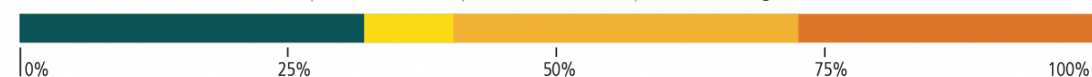
Code shared

Open | Yes | Some info | No | N/A



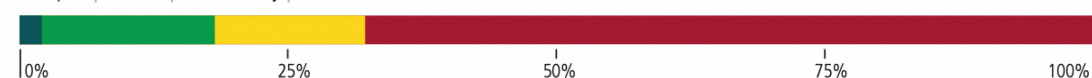
Analysis reported

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Data shared

Open | Raw | Summary | No



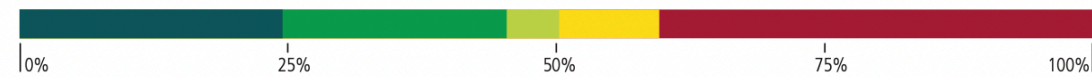
Challenges

DESIGNED
193 experiments

All needed clarifications with 49%
few/some; 20% strong/extreme

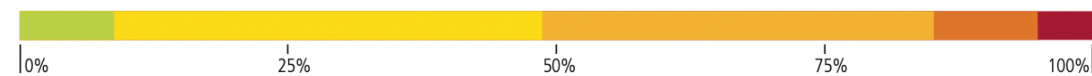
Authors helped

Extreme | Very | Moderate | Some | Minimal | No



Protocol clarifications needed

Few | Some | Moderate | Strong | Extreme



Reagents offered

Yes | No | N/A



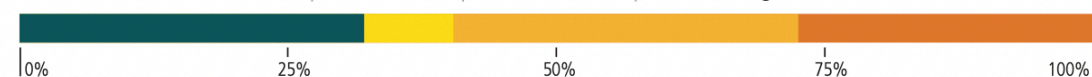
Code shared

Open | Yes | Some info | No | N/A



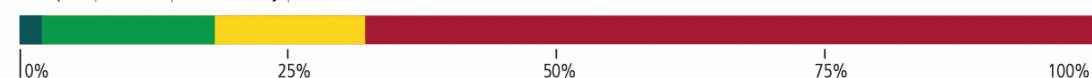
Analysis reported

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Data shared

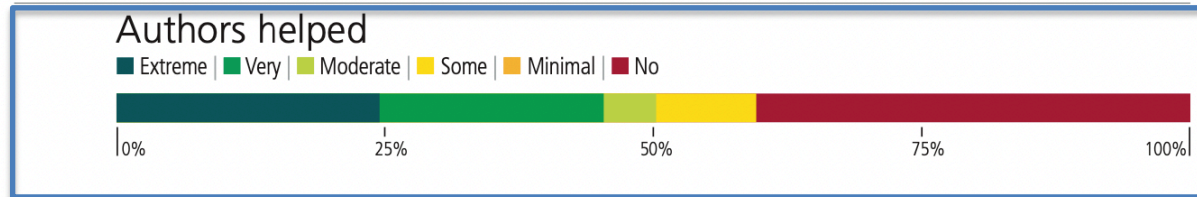
Open | Raw | Summary | No



Challenges

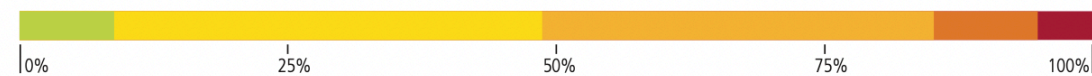
DESIGNED
193 experiments

41% extremely/very helpful, 32% not at all helpful/no response



Protocol clarifications needed

■ Few ■ Some ■ Moderate ■ Strong ■ Extreme



Reagents offered

■ Yes ■ No ■ N/A



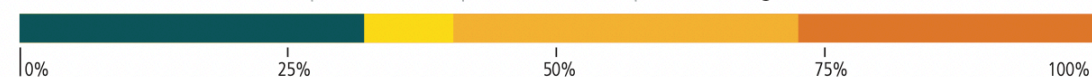
Code shared

■ Open ■ Yes ■ Some info ■ No ■ N/A



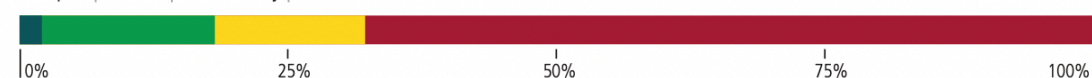
Analysis reported

Statistical inference: ■ Test known ■ Test unknown ■ No, but variation ■ No, but image

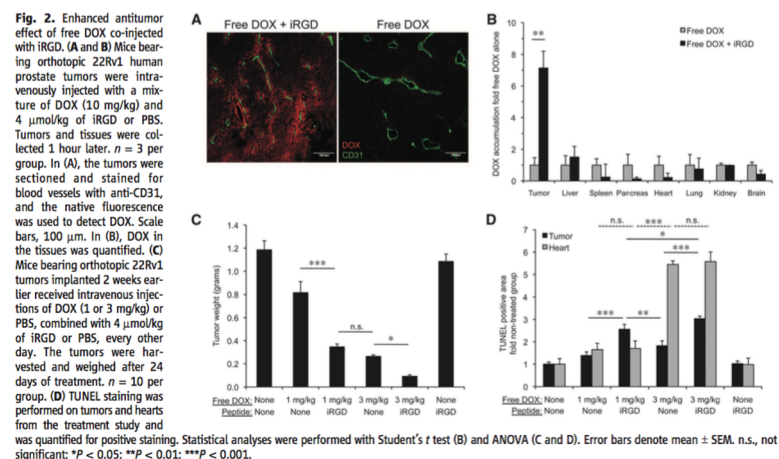


Data shared

■ Open ■ Raw ■ Summary ■ No



From the publication



Materials and Methods

In vivo skin permeability assay was performed as described elsewhere (6).

Immunofluorescence. Tissue preparation and staining of the cryosections were performed as described (3). The primary antibodies were rat anti-mouse CD31 (BD Biosciences), FITC-labeled HLA-A,B,C (BD Biosciences), and FITC-labeled H-2kd (BD Biosciences) monoclonal, and rabbit anti-T7 phage polyclonal (6) antibodies. The secondary antibodies, Alexa Fluor 594 goat anti-rat, 647 goat anti-rat, and 488 donkey anti-rabbit antibodies were from Molecular Probes.

Immunohistochemistry. Tissue preparation and staining of the cryosections were performed as described (3). The primary antibodies used were biotinylated rabbit anti-FITC/Oregon green polyclonal (Molecular Probes), and biotinylated mouse anti-dextran monoclonal (Stemcell Technologies, Vancouver, BC, Canada) and biotinylated rat anti-mouse CD31 monoclonal (BD Biosciences) antibodies. Biotinylated secondary polyclonal antibodies were goat anti-rabbit and rabbit anti-human (both from Pierce Biotechnology, Rockford, IL). In some experiments, tissue sections were stained with a TUNEL assay kit (In Situ Cell Death Detection Kit, POD; Roche Applied Science, Indianapolis, IN), and quantified for the positive areas with a scanner as described elsewhere in this manuscript.

Ex vivo tumor penetration assay. PPC-1 subcutaneous tumors (about 1 cm in diameter) were excised and placed in DMEM containing 1% BSA. The tumors were first incubated with the inhibitors or peptides for 20 min at 4°C. G₇ or iRGD phage were then added to the solution and the tumors were further incubated for 90 min at 37°C or 4°C. The tumors were then washed with cold DMEM containing 1% BSA, fixed in 4% paraformaldehyde, sectioned, immunofluorescently stained, and viewed under a confocal microscope.

Quantification of ABX in tumors and tissues was performed as described elsewhere (3).

Open Science Framework

Registered Report Additional Materials

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Registered Report Additional ...

OSF Storage

- Study 9_SF9A.pzfx
- Study_15_estimated_dat...
- Study_15_Figure_2B_gra...
- Study_15_Figure_2B_two...
- Study_15_Figure_2C_gra...
- Study_15_Figure_2D_gra...
- Study_15_power_calculat...
- Study_15_Protocol_4_Fig...
- Study_15_Protocol_4_Fig...
- Study_15_Protocol_4_Fig...
- Study_15_Protocol_5_Fig...
- Study_15_registered_rep...

igs

add a tag

Page: 4 of 9

Automatic Zoom

Test conditions:

- Orthotopic tumor bearing mice treated with Dox in combination with iRGD

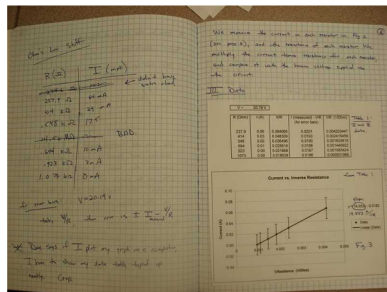
Steps:

- Generate tumor bearing mice as per Protocol 1.
- Allow tumors to grow for two weeks.
- Inject mice with drugs in combination:
 - Negative control: Inject mice intravenously with 10 mg/kg Doxorubicin (Dox) (Doxorubicin hydrochloride, Sigma-Aldrich Cat #????) (suspended in 100 μ l PBS) and with 100 μ l PBS.
 - Can the authors please provide the catalog number of the Dox purchased from Sigma?
 - Negative control: Inject mice intravenously with 10 mg/kg Dox (suspended in 100 μ l PBS) and (?) RDGfK (suspended in 100 μ l PBS).
 - Experimental: Inject mice intravenously with 10 mg/kg Dox (suspended in 100 μ l PBS) and 4 μ mol/kg iRGD (suspended in 100 μ l PBS).
- One hour later, sacrifice mice and excise tumors:
 - Euthanize mice
 - How were mice euthanized? CO₂ and cervical dislocation?
 - Perfuse with PBS + 1% BSA.
 - Was the PBS + 1% BSA cold? Perfusion was accomplished how - through the left ventricle of the heart?
 - Excise prostate tumor tissue.
- Process, embed and section tumor tissue
 - Were tumors fixed? If so, in what fixative and for how long? Given that Dox native fluorescence is used later in the protocol, fixation method can affect the signal from the free Dox.
 - Embed tumor tissue in Optimal Cutting Temperature medium (OCT) (purchased from where?) and freeze.
 - How thick were the sections cut from the tumors?
 - Materials and Methods section "Immunofluorescence" and "Immunohistochemistry" state: "Tissue preparation and staining of the cryosections were performed as described (3)". There is no reference 3 in the Supplemental Materials (only References 35-37), while Reference 3 in the main article is a listing of amino acid abbreviations. Can the authors please provide the details of tissue processing and staining?
- Immunostain tumor tissue with rat anti-CD31 (BD Biosciences, ?). Use native fluorescence to image Dox.
 - Can the authors please provide the Catalog number of the anti-CD31 antibodies?

From the publication

- What antibodies were used?
- What sex were the mice?
- How long was the treatment period for?
- What instrument/software was used?
- What was the staining protocol used?
- How was positive area defined and measured?
- Can you share the data with us?

From the authors



“I think that nuance was lost trying to edit the text down to size.”

“I do not have those data with me and will need to dig a bit in my back-ups”



“we have been working with the protocol document. Problem was that different experiments were done by different authors”

“I do not have the raw data anymore”



“some of the details you are seeking are not readily accessible”

Barriers to Conducting Replications in Experiments

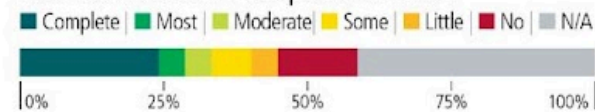
By research stage

COMPLETED

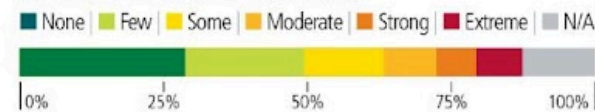
50 experiments

BARRIERS

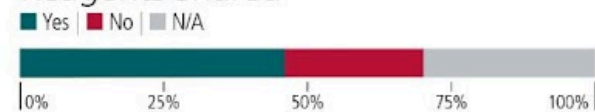
Modifications implemented



Modifications needed



Reagents shared



CONDUCTED

87 experiments

Barriers to Conducting Replications in Experiments

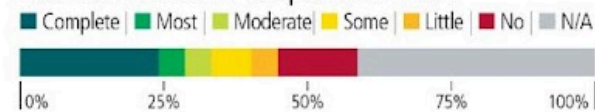
By research stage

COMPLETED

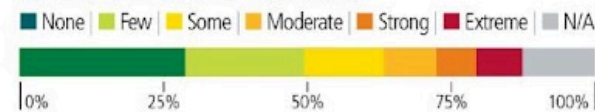
50 experiments

BARRIERS

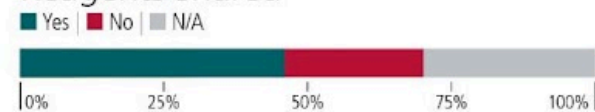
Modifications implemented



Modifications needed



Reagents shared



66% actually shared reagents

CONDUCTED

87 experiments

Barriers to Conducting Replications in Experiments

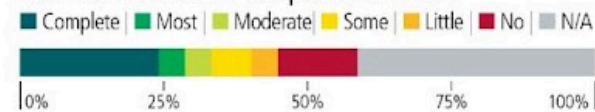
By research stage

COMPLETED

50 experiments

BARRIERS

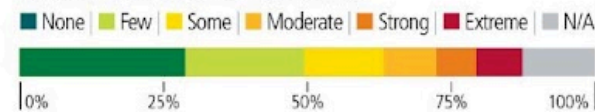
Modifications implemented



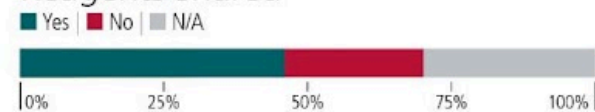
CONDUCTED

87 experiments

Modifications needed



Reagents shared



67% required modifications with 39% few/some; 17% strong/extreme

Barriers to Conducting Replications in Experiments

By research stage

COMPLETED

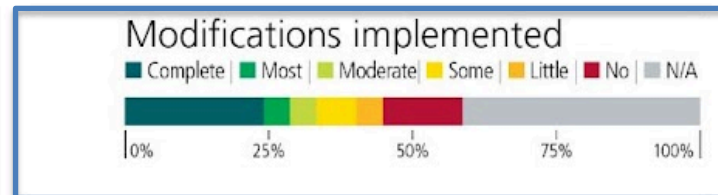
50 experiments

BARRIERS

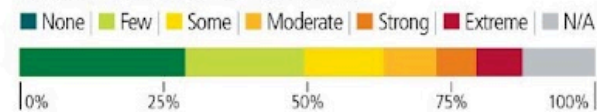


CONDUCTED

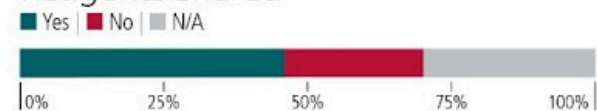
87 experiments



Modifications needed



Reagents shared



41% completely implemented

Testing replicability impeded by...

- Lack of transparency and accessibility of methodology
- Lack of sharing of original data and reagents
- Lack of communication for obtaining needed information
- Unexpected challenges with protocols during experimentation
- Resource challenges (cost, time, uncertainty) inflated by all the above



FEATURE ARTICLE



RESEARCH ARTICLE



REPRODUCIBILITY IN CANCER BIOLOGY

Challenges for assessing replicability in preclinical cancer biology

Abstract We conducted the [Reproducibility Project: Cancer Biology](#) to investigate the replicability of preclinical research in cancer biology. The initial aim of the project was to repeat 193 experiments from 53 high-impact papers, using an approach in which the experimental protocols and plans for data analysis had to be peer reviewed and accepted for publication before experimental work could begin. However, the various barriers and challenges we encountered while designing and conducting the experiments meant that we were only able to repeat 50 experiments from 23 papers. Here we report these barriers and challenges. First, many original papers failed to report key descriptive and inferential statistics: the data needed to compute effect sizes and conduct power analyses was publicly accessible for just 4 of 193 experiments. Moreover, despite contacting the authors of the original papers, we were unable to obtain these data for 68% of the experiments. Second, none of the 193 experiments were described in sufficient detail in the original paper to enable us to design protocols to repeat the experiments, so we had to seek clarifications from the original authors. While authors were *extremely or very helpful* for 41% of experiments, they were *minimally helpful* for 9% of experiments, and *not at all helpful* (or did not respond to us) for 32% of experiments. Third, once experimental work started, 67% of the peer-reviewed protocols required modifications to complete the research and just 41% of those modifications could be implemented. Cumulatively, these three factors limited the number of experiments that could be repeated. This experience draws attention to a basic and fundamental concern about replication – it is hard to assess whether reported findings are credible.

TIMOTHY M ERRINGTON*, ALEXANDRIA DENIS†, NICOLE PERFITO‡, ELIZABETH IORNS AND BRIAN A NOSEK

Investigating the replicability of preclinical cancer biology

Timothy M Errington^{1*}, Maya Mathur², Courtney K Soderberg¹, Alexandria Denis^{1†}, Nicole Perfito^{1‡}, Elizabeth Iorns³, Brian A Nosek^{1,4}

¹Center for Open Science, Charlottesville, United States; ²Quantitative Sciences Unit, Stanford University, Stanford, United States; ³Science Exchange, Palo Alto, United States; ⁴University of Virginia, Charlottesville, United States

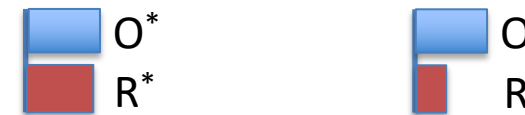
Abstract Replicability is an important feature of scientific research, but aspects of contemporary research culture, such as an emphasis on novelty, can make replicability seem less important than it should be. The [Reproducibility Project: Cancer Biology](#) was set up to provide evidence about the replicability of preclinical research in cancer biology by repeating selected experiments from high-impact papers. A total of 50 experiments from 23 papers were repeated, generating data about the replicability of a total of 158 effects. Most of the original effects were positive effects (136), with the rest being null effects (22). A majority of the original effect sizes were reported as numerical values (117), with the rest being reported as representative images (41). We employed seven methods to assess replicability, and some of these methods were not suitable for all the effects in our sample. One method compared effect sizes: for positive effects, the median effect size in the replications was 85% smaller than the median effect size in the original experiments, and 92% of replication effect sizes were smaller than the original. The other methods were binary – the replication was either a success or a failure – and five of these methods could be used to assess both positive and null effects when effect sizes were reported as numerical values. For positive effects, 40% of replications (39/97) succeeded according to three or more of these five methods, and for null effects 80% of replications (12/15) were successful on this basis; combining positive and null effects, the success rate was 46% (51/112). A successful replication does not definitively confirm an original finding or its theoretical interpretation. Equally, a failure to replicate does not disconfirm a finding, but it does suggest that additional investigation is needed to establish its reliability.

How to assess replicability?

- Same direction



- Same direction and statistically significant



- Original effect size in replication confidence interval



- Replication effect size in original confidence interval



- Replication effect size in prediction interval



- Replication effect size compared to original effect size



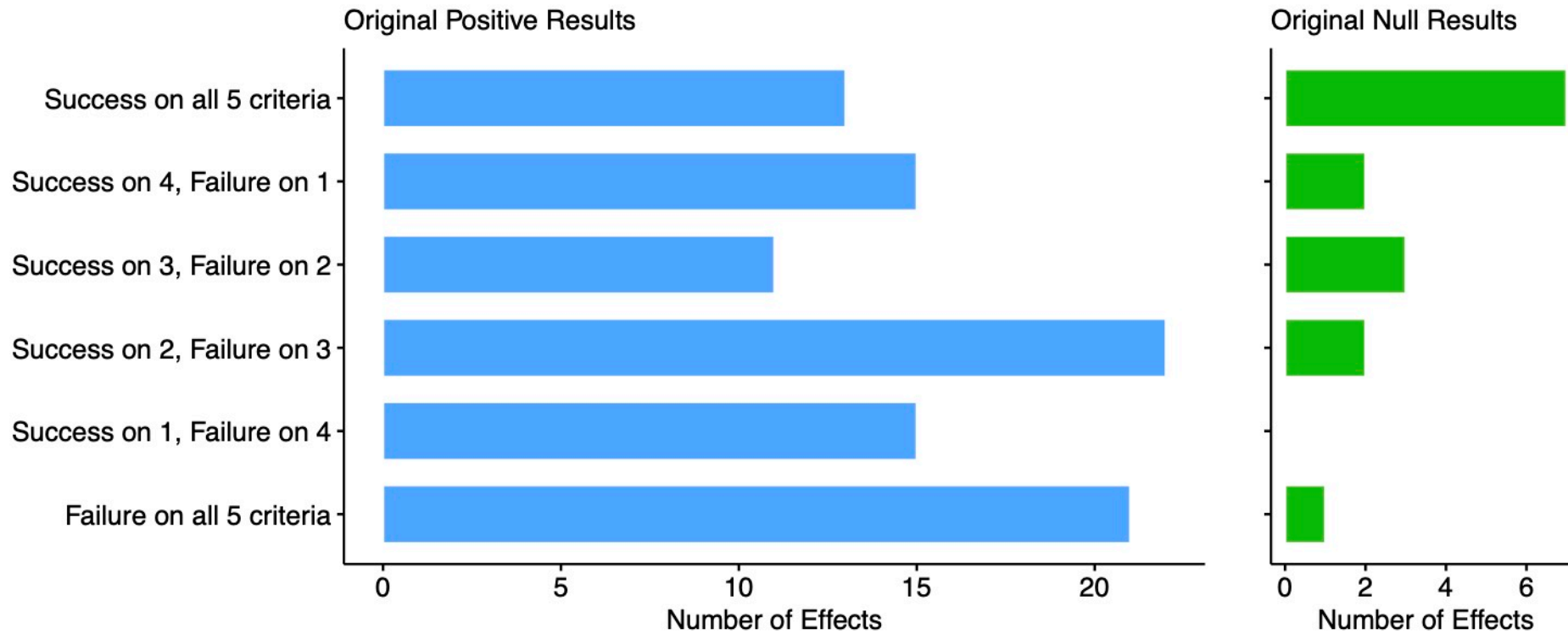
- Direction and statistical significance of meta-analysis



Summarizing across five dichotomous replication success criteria

- 47%: Same direction and statistically significant
- 25%: Original effect size in the replication 95% confidence interval
- 48%: Replication effect size in the original 95% confidence interval
- 61%: Replication effect size in the 95% prediction interval
- 63%: Meta-analysis of original and replication

Outcomes

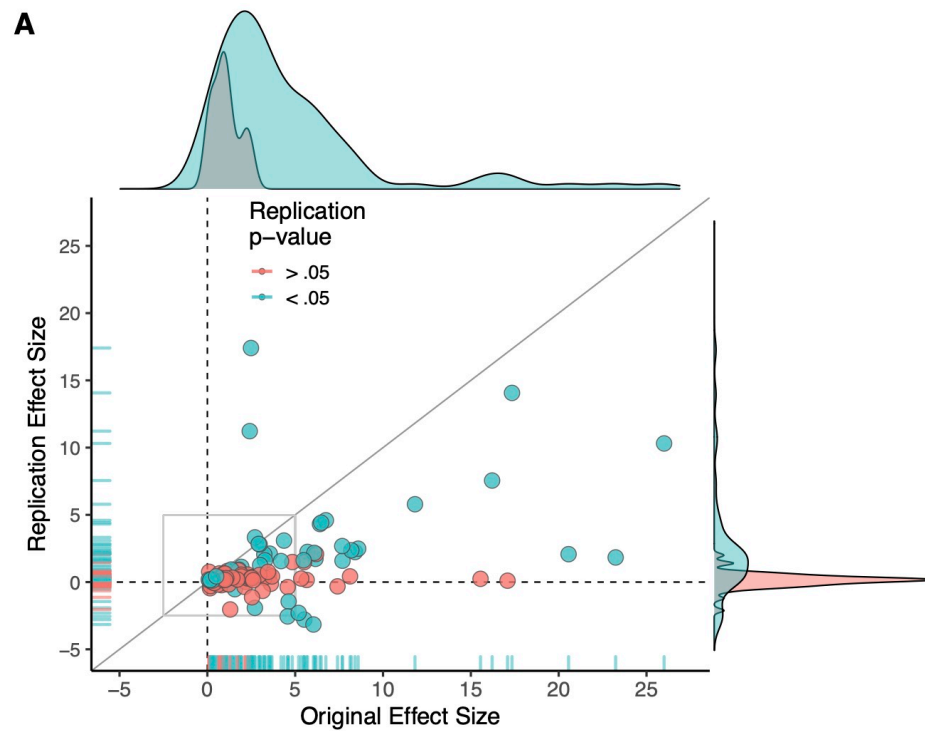


46% succeeded on most criteria, 54% failed on most criteria

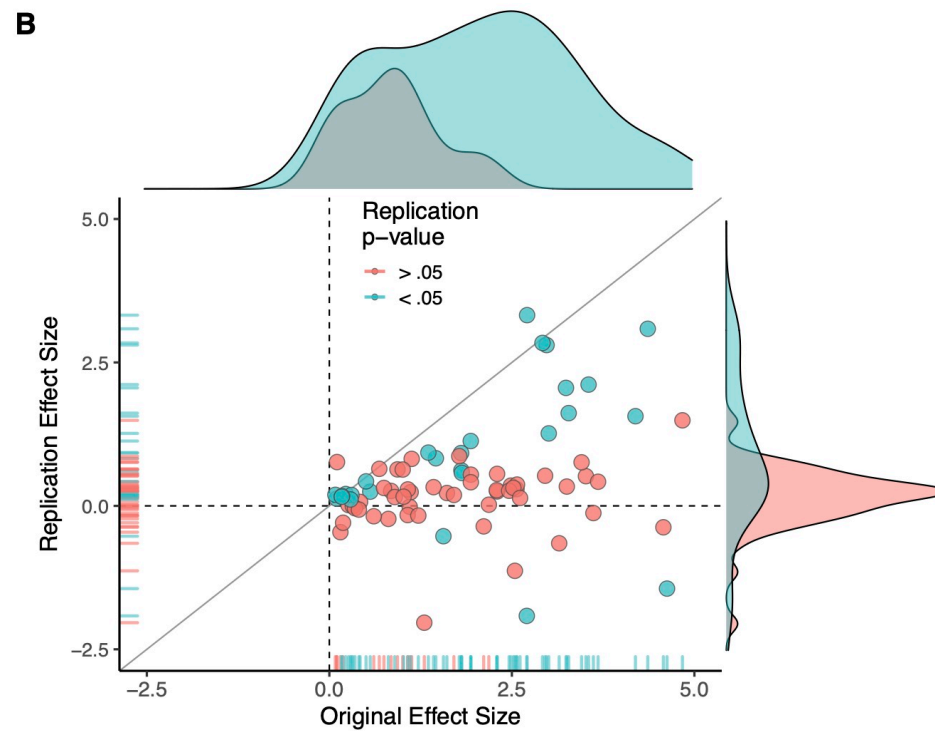
80% of original null effects succeeded on most criteria

40% of original positive effects succeeded on most criteria

Replication effects compared with original effects

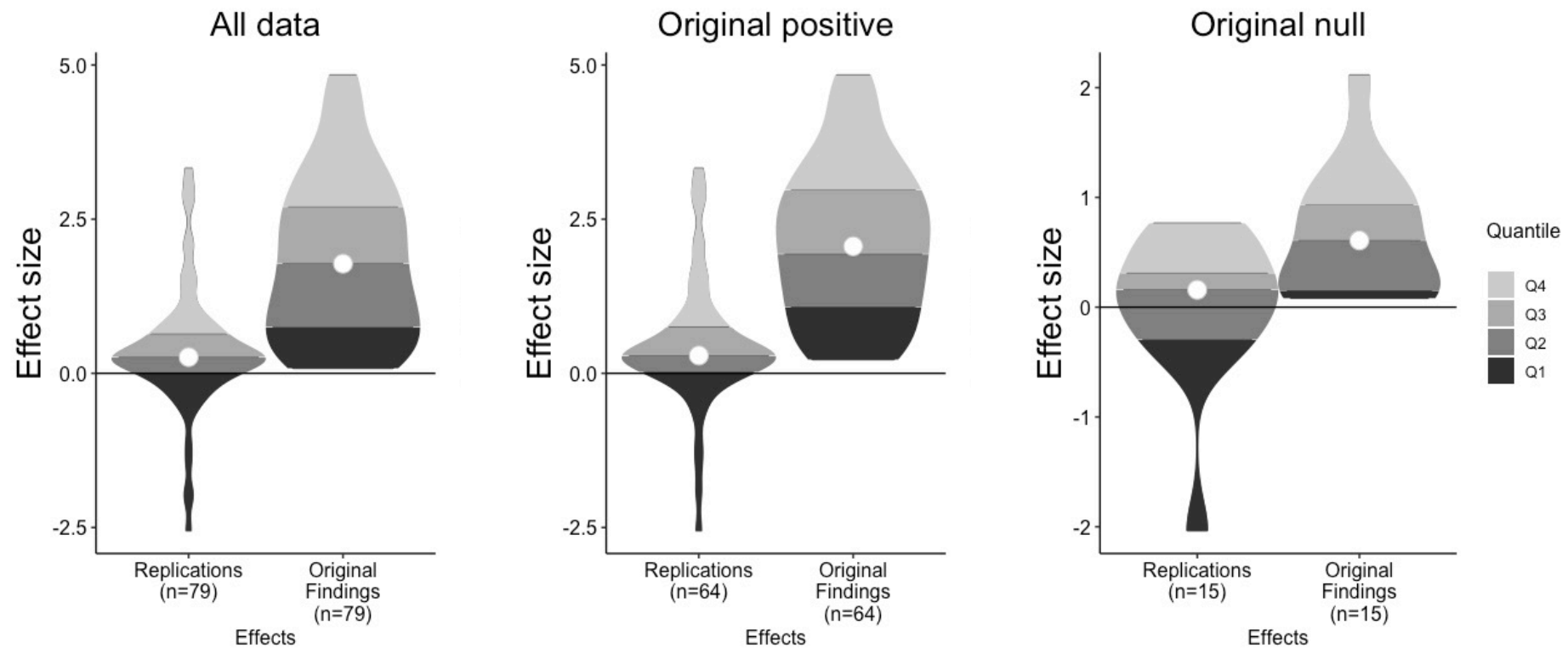


Replications 85% smaller on average



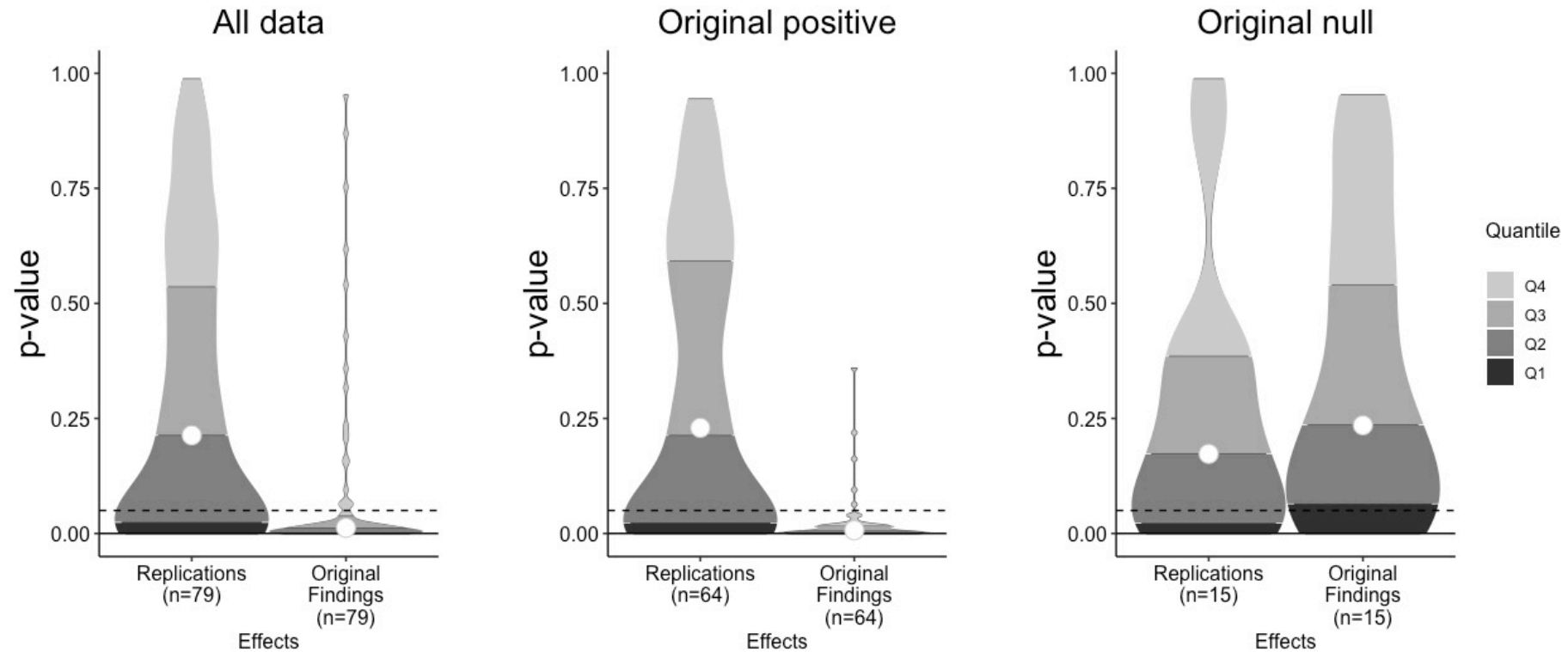
Zoomed in on effect sizes <5

Replication effects compared with original effects



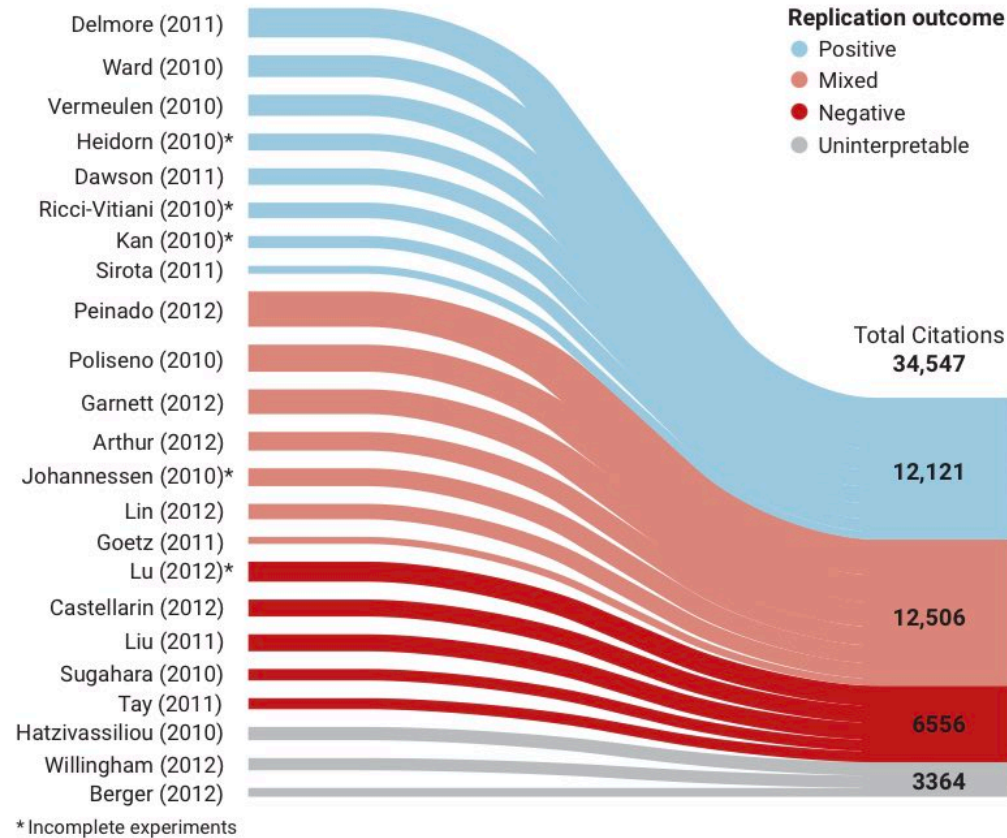
effect sizes <5

Replication p-values compared with original p-values



effect sizes <5

Subjective assessment



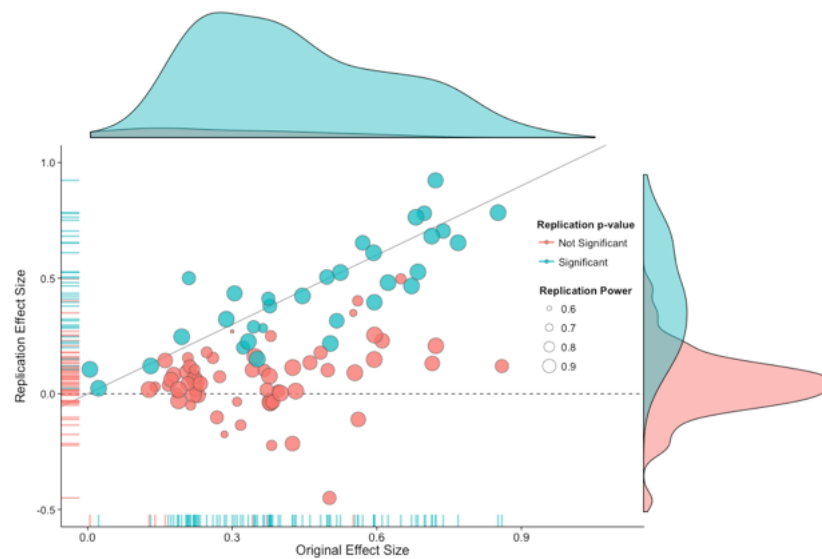
GRAPHIC: K. FRANKLIN/SCIENCE; DATA: REPRODUCIBILITY PROJECT: CANCER BIOLOGY

Meta-analysis conclusions

- Replication effects were much weaker than originals
- “Success” was low across replication criteria with variability due, in part to liberalness of the test
- Positive results were half as likely to replicate as null results
- Animal and non-animal declines similar magnitudes - animal effects lower success rate because small original effect sizes
- There is room for improvement

Replication in...

Psychology

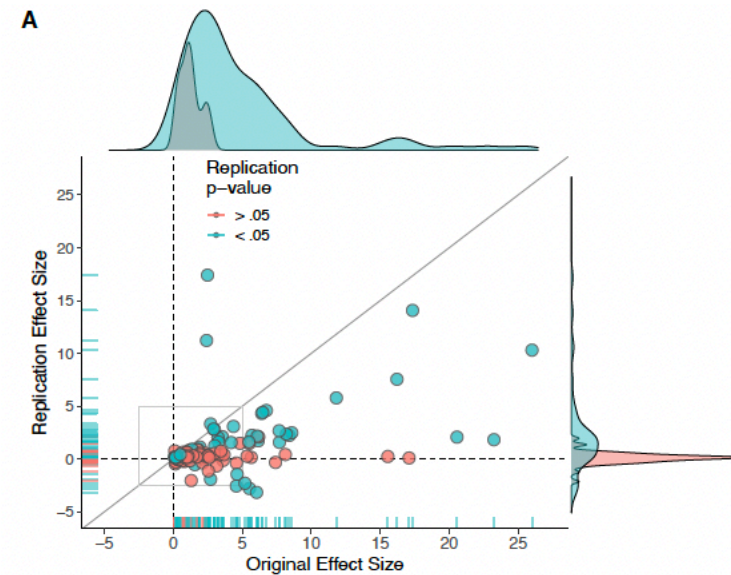


Open Science Collaboration, 2015 *Science*

Replications

*36% significant, same direction as original
50% smaller than original on average*

Cancer Biology



Errington et al., 2021 *eLife*

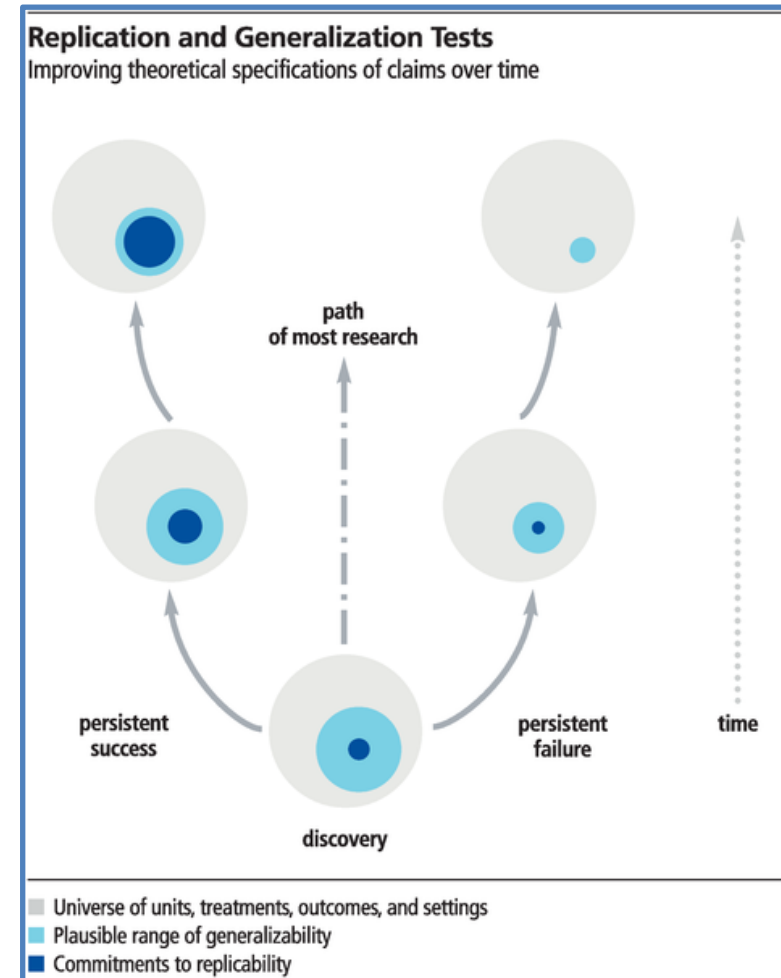
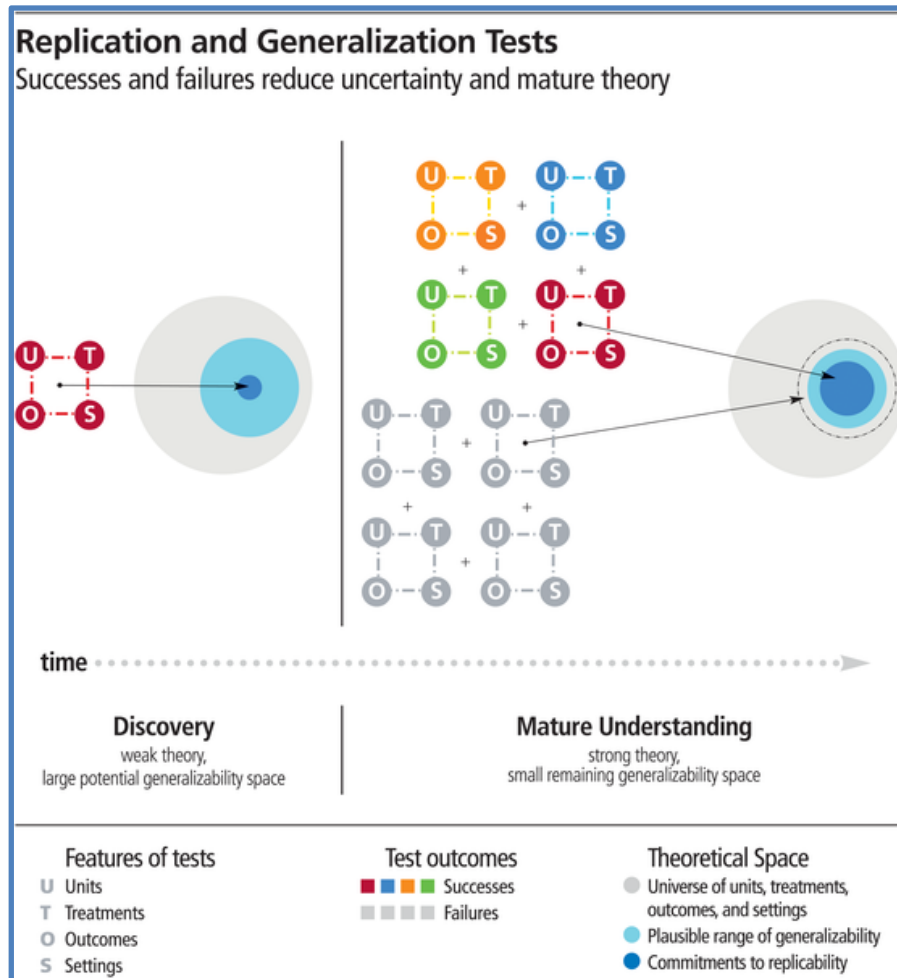
Replications

*43% significant, same direction as original
85% smaller than original on average*

Interpreting failures to replicate

- A failure to replicate could mean:
 - The original finding was a false positive
 - The replication was a false negative
 - Both are “true” and key conditions in the experimental design differ

Do we know the conditions necessary to observe a finding?

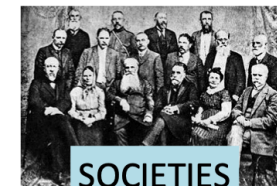


What can we do?

- Incentivize open science practices in your community
 - Aligning institutional policies with open science practices (e.g., [NASEM Roundtable](#))
 - Journal polices that incentivize open practices (e.g., [TOP Guidelines](#))
 - Assessment of researchers and scholarly research (e.g., [DORA](#))
 - Training on reproducible and open science practices

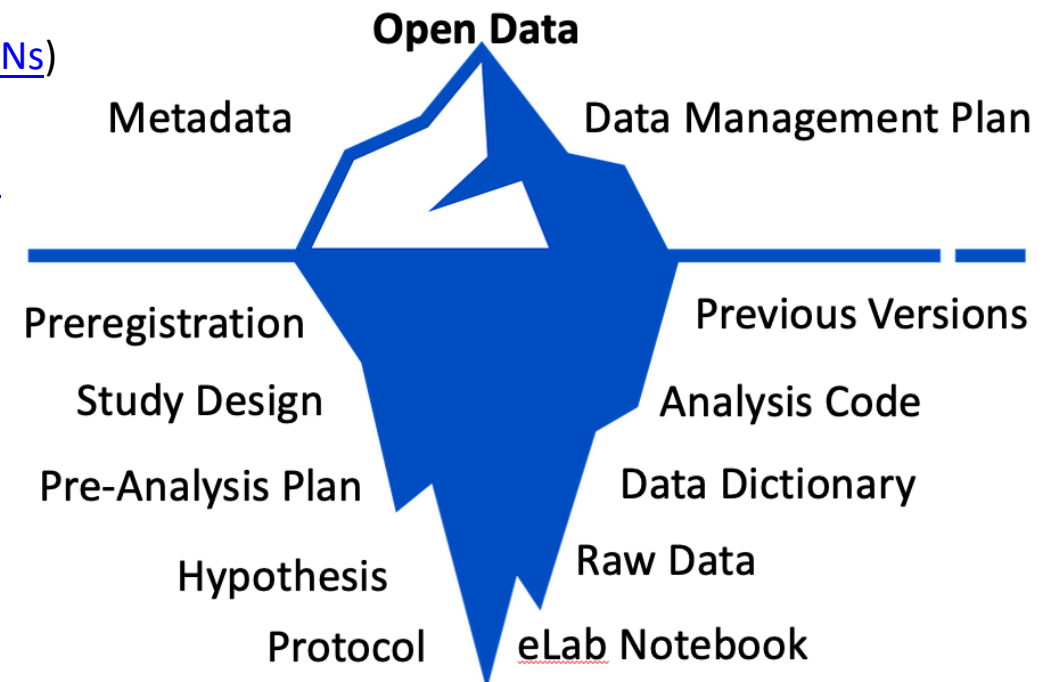


social system



What can we do?

- Incorporate open science practices in your research
 - Share data/code/etc using repositories (e.g., [NIH GREI Repositories](#))
 - Deposit reagents in repositories (e.g., [addgene](#))
 - Make detailed protocols open (e.g., [ELNs](#))
 - Try [preregistration/Registered Reports](#)



Study 15: Replication of Sugahara et al., 2010 (Science)

Contributors: Christine Mantis, Irawati Kandela, Fraser Aird, [Elizabeth Iorns](#), [Alexandria Denis](#), [Stephen Williams](#), [Nicole Perfito](#), [Tim Errington](#)

Affiliated institutions: [Center For Open Science](#), [Laura and John Arnold Foundation](#)

Date created: 2013-10-22 04:11 PM | Last Updated: 2017-01-18 02:44 PM

Identifiers: DOI 10.17605/OSF.IO/XU1G2 | ARK c7605/osf.io/xu1g2

Category: Project

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Wiki

Replication Study: Coadministration of a Tumor-Penetrating Peptide Enhances the Efficacy of Cancer Drugs

Abstract

In 2015, as part of the [Reproducibility Project: Cancer Biology](#), we published a Registered Report ([Kandela et al., 2015](#)), that described how we intended to replicate selected experiments from the paper "Coadministration of a tumor-penetrating peptide enhances the efficacy of cancer d...

[Read More](#)

Files

Name	Modified
Study 15: Replication of Sugahara et al., 2010 (Science)	
OSF Storage	
Replication_Study_15.docx	2016-10-25 05:39 PM
Replication_Study_15.pdf	2016-10-25 05:39 PM
Replication_Study_15.Rmd	2017-01-11 01:42 PM

Citation

osf.io/xu1g2

Components

Quantifying the amount of Dox present in tumor tissue and major organs in mice treated with Dox with or without iRGD

Mantis, Kandela, Aird & 5 more
39 contributions

Effect of Dox alone or Dox in combination with iRGD on tumor growth and total body weight

Mantis, Kandela, Aird & 5 more
47 contributions

Assessment of TUNEL staining of tumor and heart tissue after treatment

Mantis, Kandela, Aird & 5 more
39 contributions

Meta-analyses

Iorns, Denis, Williams & 2 more
117 contributions

iRGD peptide synthesis

Chou
63 contributions

Dose administration

Mice were intravenously injected (IV) by tail vein, based on body weight on injection day. For tumor and organ penetrance analysis, [Figure 1](#), mice were anesthetized, perfused, and sacrificed 1 hr after drug administration. For multi-day experiments mice were injected by IV every other day for 24 days based on body weight on injection day. On the last day, mice were anesthetized, perfused, and sacrificed 1 hr after drug administration. Further details of these methods are available at (<https://osf.io/bkhnp/>).

Dose administration

Mice were intravenously injected (IV) by tail vein, based on body weight on injection day. For tumor and organ penetrance analysis, [Figure 1](#), mice were anesthetized, perfused, and sacrificed 1 hr after drug administration. For multi-day experiments mice were injected by IV every other day for 24 days based on body weight on injection day. On the last day, mice were anesthetized, perfused, and sacrificed 1 hr after drug administration. Further details of these methods are available at [\(https://osf.io/bkhnp/\)](https://osf.io/bkhnp/)

Dose administration

Mice were intravenously injected (IV) by tail vein, based on body weight on injection day. For tumor and organ penetrance analysis, Figure 1, mice were

TUNEL analysis

TUNEL (EMD Millipore, cat # S7100) stained slides were evaluated by Dr. Gennadiy Bondarenko and Dr. Andrey Ugolkov. Images were captured using a Carl Zeiss Axial Lab A1 microscope and a 40x objective, by Dr. Ugolkov, or a Olympus BX45 microscope and a 40x objective, by Dr. Bondarenko (images available at: <https://osf.io/3fs27/>). Drs. Bondarenko and Ugolkov were blinded to group allocation, only receiving the animal ID with H or P designation to indicate heart or prostate tumor. The frequency of apoptosis was calculated as an apoptotic index, in which the proportion of cells undergoing apoptosis was expressed as a percentage of all cells observed. The apoptotic index of each tissue sample was calculated as the number of TUNEL-positive cells and bodies per 500 cells/microscopic view or 2500 cells/slide (5 slides/tissue), counted in five randomly selected microscopic fields in each tissue sample. Percent apoptotic index was calculated with the following formulation: $(i/500) \times 100\%$. i = cell undergoing apoptosis. An average was taken of the apoptotic index from all five fields of all five slices (25 fields total) which is considered one biological replicate. Negative and positive control sections were stained in parallel to the tumor and heart samples and are available at (<https://osf.io/gmcyt/>). Original counts are available at <https://osf.io/pbg7x/>.

Dose administration

Mice were intravenously injected (IV) by tail vein, based on body weight on injection day. For tumor an

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TUNEL analysis

TUNEL (EMD Millipore, c
Bondarenko and Dr. Andr
A1 microscope and a 40x
40x objective, by Dr. Bonc
Bondarenko and Ugolkov
with H or P designation to
was calculated as an apop
apoptosis was expressed a
tissue sample was calculat
cells/microscopic view or
selected microscopic field
with the following formul
was taken of the apoptotic
is considered one biologic
in parallel to the tumor an
Original counts are availa

Statistical analysis

Statistical analysis was performed with R software (RRID: [SCR_001905](https://scicr.org/RRID/SCR_001905)), version 3.2.2 (R Core Team, 2016). All data csv files and analysis scripts are available at (<https://osf.io/xu1g2/>). Confirmatory statistical analysis was pre-registered (<https://osf.io/9hr2d/>) before the experimental work began as outlined in the Registered Report (Kandela et al., 2015). Additional exploratory analysis (area under the curve) was performed using the weights of the mice over the treatment period. Data were checked to ensure assumptions of statistical tests were met. A meta-analysis of a common original and replication effect size was performed using a random effects model and the *metafor* R package (Viechtbauer, 2010). (available at <https://osf.io/ymxaz/>). The original study data were extracted *a priori* from the published figures by determining the mean and upper/lower error values for each data point. The extracted data were published in the Registered Report (Kandela et al., 2015) and were used in the power calculations to determine the sample sizes for this study. In the meta-analyses where Glass' Δ was used, because of unequal variance between the two conditions being compared, the standard deviation of DOX + PBS was used in the calculations.

Conclusions



Figure 4.

TUNEL staining of mouse tissues.

Mice bearing orthotopic 22Rv1 human prostate tumors were intravenously injected with PBS, 1 mg/kg DOX and PBS (DOX + PBS), or 1 mg/kg DOX and 4 μ mol/kg of ... (DOX + iRGD). TUNEL staining was performed on tumor and heart sections of each animal. (A) Boxplot of mean apoptotic index calculated from TUNEL stained tumor sections. TUNEL scores were normalized to the average score of tumors from PBS treated mice. Means reported and error bars represent s.e.m. Number of mice per condition ($n=6$; $n=18$ mice total). One-way ANOVA on apoptotic index of all groups; $F(2, 15) = 1.378$, $p=0.282$. Planned contrast between DOX + PBS and DOX + iRGD; $t(15) = 0.435$, $p=0.670$ with *a priori* alpha level = 0.05. Representative images of TUNEL staining of tumor sections from PBS (B), DOX + PBS (C), or DOX + iRGD (D) treated mice. Representative images of TUNEL staining of heart sections from PBS (E), DOX + PBS (F), or DOX + iRGD (G) treated mice. Additional details for this experiment can be found at <https://osf.io/7eynw/>.

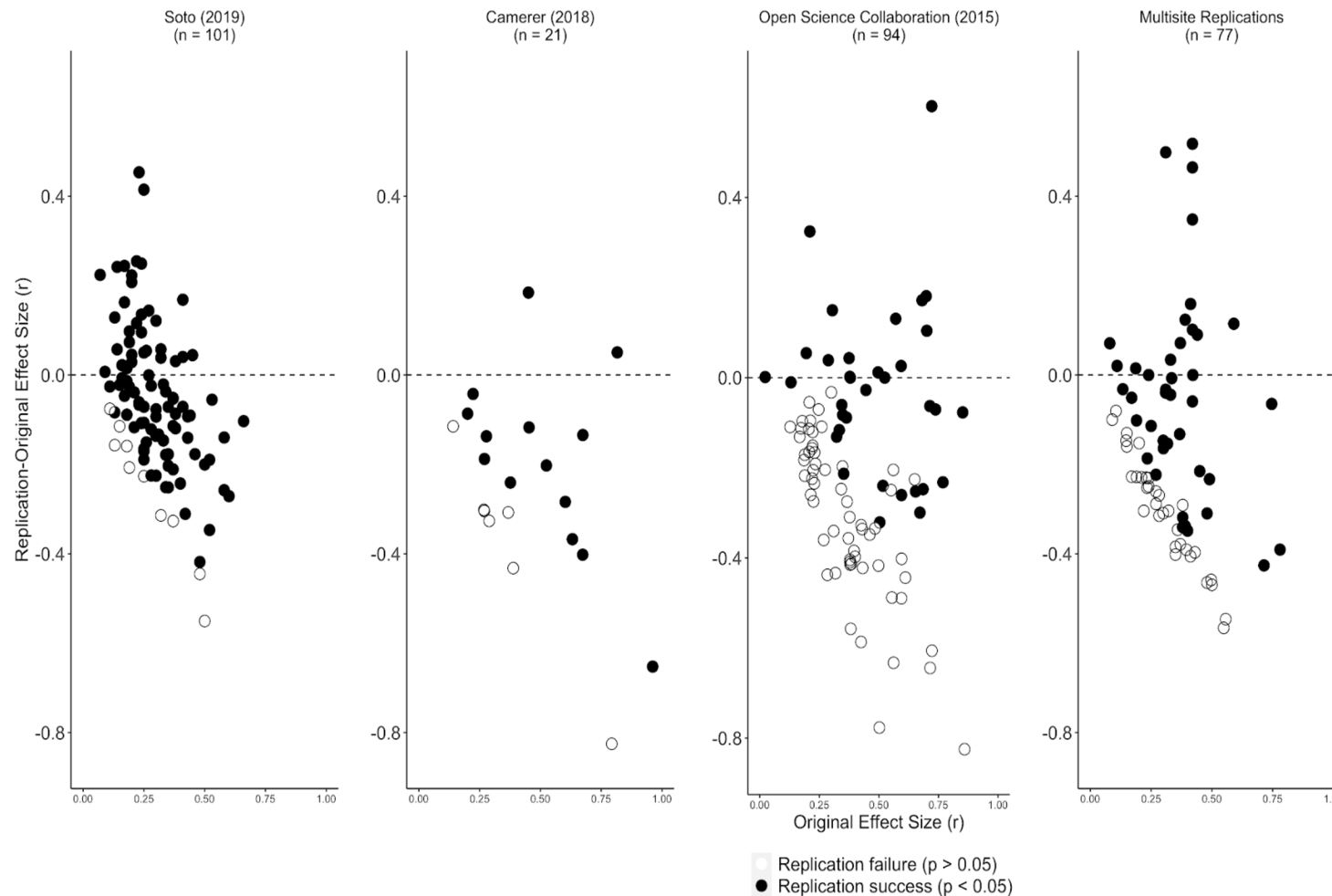
All RP:CB papers point to the OSF components which host not only the selected representative images, but all images collected, methods, analyzes, and figure generation.

“ An article about **computational** science in a scientific publication is not the scholarship itself, it is merely advertising of the scholarship. The actual scholarship is the complete **software development environment**, and the complete set of instructions which generated the figures. ”

Buckheit & Donoho, 1995

Wavelab and Reproducible Research

Do adopting open science practices increase replicability?



**Weaker
evidence than
original study**



More information

{Take a picture}

- These slides: <https://osf.io/yf259>
- Center for Open Science: <https://cos.io/>
- Reproducibility Project: Cancer Biology: <https://cos.io/rpcb>
- NASEM Roundtable: <https://www.nationalacademies.org/our-work/roundtable-on-aligning-incentives-for-open-science>
- TOP Guidelines: <https://www.cos.io/top/>
- DORA: <https://sfdora.org/>
- NIH GREI: <https://datascience.nih.gov/news/nih-office-of-data-science-strategy-announces-new-initiative-to-improve-data-access>
- OSF: <https://osf.io/>
- Preregistration: <https://cos.io/prereg/>
- Registered Reports: <https://cos.io/rr/>
- Addgene: <https://www.addgene.org/>
- Resources on ELNs: <https://datamanagement.hms.harvard.edu/analyze/electronic-lab-notebooks>

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